

Catalytic Stereodivergent Synthesis of Steroid-Fulleropyrrolidine Hybrids

Margarita Suárez,^{†*} Alberto Ruiz,[†] Luis Almagro,[†] Julieta Coro,[†] Enrique E. Maroto,[§] Salvatore Filipone,[§] Dolores Molero,[±] Roberto Martínez-Álvarez,[§] Nazario Martín^{§*}

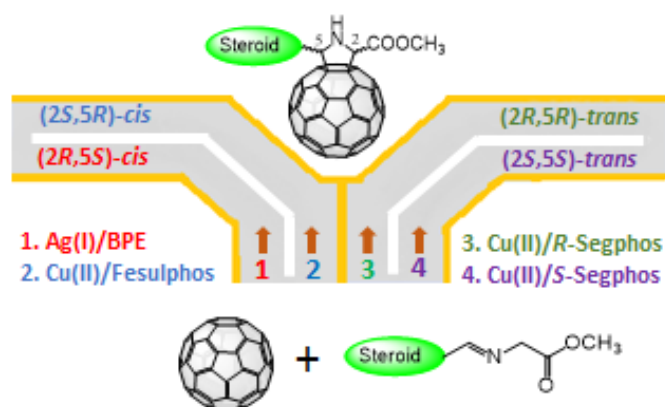
[†]Laboratorio de Síntesis Orgánica, Facultad de Química, Universidad de la Habana, 10400 La Habana, Cuba.

[§]Departamento de Química Orgánica I, Facultad de Ciencias Químicas, Universidad Complutense de Madrid, 28040 Madrid, Spain.

[±]CAI RMN Universidad Complutense de Madrid, 28040 Madrid, Spain.

E-mail: msuarez@fq.uh.cu, nazmar@quim.ucm.es

This work is dedicated to the memory of Prof. Dr. José L. Mola Gárate



ABSTRACT

The diastereoselective synthesis of *cis* and *trans* steroid-fulleropyrrolidines hybrids by reaction of *N*-metalated azomethine ylides [Cu(II) or Ag(I)] with the appropriate chiral ligand and C₆₀ is described. The experimental findings reveal that the azomethine ylide stabilized by an allylic group cycloadds to [60]fullerene in an efficient manner and with a good diastereomeric excess. Furthermore, the new generated stereocenters are fully controlled by the catalytic systems used without being influenced by the chirality of the steroid. Interestingly, by this synthetic methodology the each one of the four possible stereoisomers have efficiently been obtained and characterized by CD spectra.

INTRODUCTION

The use of fullerenes for bio-medical applications is still an underdeveloped research area.¹ Therefore, synthetic approaches towards fullerenes covalently bonded to natural

products are of interest.² On the other hand, it is well known that steroids are biologically relevant molecules showing bio-medical applications.^{3,4} In this regard, conjugating steroids to fullerenes is a topic that has not been properly addressed so far that can lead to interesting molecules with different biological properties.⁵⁻¹¹

In previous work we have carried out the synthesis of new [60]fullerene–steroid hybrids following the Bingel-Hirsch methodology using cholesterol, β -sitosterol, and ergosterol as steroid moiety, affording a variety of methanofullerene conjugates exhibiting solubility in different organic solvents, thus obtaining promising functional chimeras with potential biomedical applications.¹² Soon afterwards, we synthesized new molecular hybrids steroid-fullerenes, using the Prato's procedure by 1,3-cycloaddition of the corresponding formyl-steroid, sarcosine and C₆₀. In that case, we have selected the naturally occurring steroid epiandrosterone, which was conveniently functionalized. This reaction gives rise to the formation of diastereoisomeric mixture of fulleropyrrolidines with poor selectivity due to the generation of a new stereogenic center in the cyclization process.¹³ Recently, we have described the multistep preparation of a fullerene hybrid dumbbell endowed with two fullerene units connected through an epiandrosterone molecule. Suitably functionalized epiandrosterone required a previous chemical modification to introduce a formyl and a malonate groups for further covalent connectivity to the C₆₀ units, first by 1,3-cycloaddition and then by a cyclopropanation reaction.¹⁴ In both cases the lack of asymmetric control on the new generated chiral centers gave stereoisomeric mixture.

On the other hand, chirality in fullerenes has recently received a lot of attention because of its importance for preparing optically active compounds for potential use in medicinal chemistry as well as in materials science.

The first successful methodology for obtaining pyrrolidinofullerenes with controlled stereochemistry was described by Martin's group in 2009 using different copper and silver salts and chiral ligands.¹⁵ This procedure paved the way to the preparation of new and versatile chiral fullerenes at will. Particularly, chiral metal complexes described proved to be versatile catalysts affording optically pure pyrrolidinofullerenes by a fully stereodivergent cycloaddition of azomethine ylides.¹⁶

Taking into account these aforementioned results, we have carried out a thorough and systematic study to expand the scope of this highly versatile stereoselective catalytic cycloaddition onto C₆₀ to other catalysts and dipoles, reporting a fully stereodivergent methodology for the synthesis of chiral pyrrolidinofullerenes by the correct choice of a

wide and easily available collection of chiral ligands, metals, and iminoesters or iminoamides.¹⁷

The aforementioned results achieved in chiral fullerenes were reported in a comprehensive review published in 2014.¹⁸ All these outcomes pave the way to obtaining enantiomerically pure fullerenes and, therefore, to their use not only in biomedical applications, where chirality is an important issue, but also in materials science, where fullerenes find their main applications.

With the aim to prepare steroid-fullerene hybrids with a controlled stereochemistry, here we report on the stereoselective 1,3-dipolar cycloaddition of steroid *N*-metalated azometine ylides to C₆₀ using a suitable combination of chiral ligand and metal salts, at low temperature, affording each one of the four possible stereoisomers in good yields and high diastereomeric excess, surpassing de values of 93%.

RESULTS AND DISCUSSION

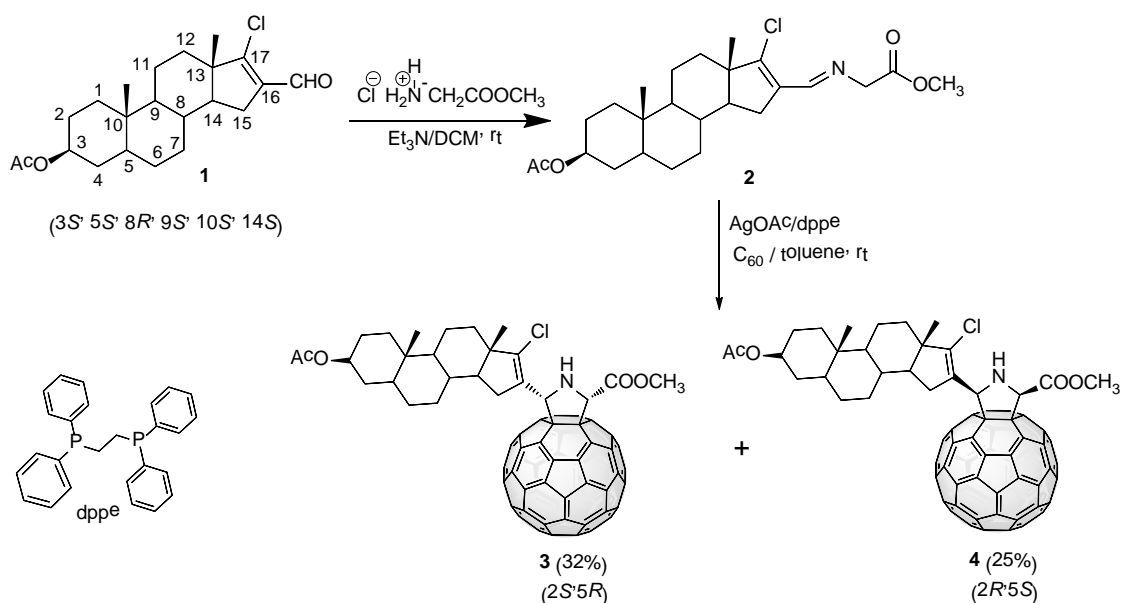
The stereoselective synthesis of steroid-fullerene hybrids from the 1,3-dipolar cycloaddition of the imine **2** onto C₆₀ has been studied by the application of metal mediated asymmetric catalysis. Since in this process the configuration of the stereogenic centers of the steroid (3'*S*, 5'*S*, 8'*R*, 9'*S*, 10'*S*, 14'*S*) does not change during the course of the reaction, a mixture of up to four optically pure stereoisomer could arise from a no controlled formation of the two new formed stereogenic carbon atoms C2 and C5: (2*R*, 5*R*), (2*S*, 5*S*), (2*S*, 5*R*) and (2*R*, 5*S*).

The synthesis of the target compounds began with the generation of the imine **2** by mixing the previously reported¹³ 3-chloro-2-formyl-17-oxo-5 α -androstan-2-ene (**1**) and glycine methyl ester hydrochloride in the presence of triethylamine using dichlorometane as solvent at room temperature (See Scheme 1). This compound was obtained as a yellow oil in 77 % yield.

The ¹H NMR spectrum of **2** shows the disappearance of the signal corresponding to the proton of the formyl group at ~10 ppm and the presence of a signal at 8.22 ppm assigned to the imine proton. In the ¹³C NMR spectrum appears signals at 170.6 ppm (O=C=O) and other at 160.7 ppm assigned to the carbon of the imine group. The structure of the compound was ascertained by mass spectrometry. The ESI-MS spectrum for compound **2** shows a peak at *m/z* = 472.3 which corresponds to [M + Na]⁺, thus supporting the proposed structure.

The use of anhydrous silver acetate along with an achiral ligand [dppe, 1,2-bis(diphenylphosphino)ethane] in toluene at room temperature limits the mixture composition to the two *cis*-diastereomers **3** and **4** as indicated by the appearance of two peaks at 6.96 and 10.71 minutes, respectively, in the HPLC chromatogram of the reaction. It is important to note, however, that the stabilization of the N-metalated azomethine ylide is provided by the allylic moiety of the steroid unit, thus resembling the benzylic one (moiety) in previous examples from literature.¹⁵

The two monoadducts were separated by flash chromatography initially with carbon disulfide, to elute unreacted C₆₀, followed by dichloromethane and dichloromethane: ethyl acetate (100:1) to obtain compounds **3** and **4**, in 32% and 25% yield, respectively, (Table 1, entry 1) see Scheme 1. These values show a modest stereoselectivity, as result of the poor chiral induction of the steroid moiety on the attack to one of the faces of the dipole in the fullerene derivative.



Scheme 1. Synthesis of *cis* diastereomers of fulleropyrrolidine-steroids hybrids.

Monoadducts **3** and **4** were fully characterized by analytical and spectroscopic techniques (see the Experimental Section and Supporting Information). ¹H NMR spectroscopy revealed the formation of compounds **3** and **4**. Besides the disappearance of the methylene protons ($\delta = 4.31$ ppm) present in **2**, the spectra of compounds **3** and **4** show the signals corresponding to the protons of the pyrrolidine ring at 5.57 ppm (H2) and 5.81 ppm (H5) for **3** and 5.58 ppm (H2) and 5.86 ppm (H5) for **4**.

The number of signals observed in the ^{13}C NMR spectra between 156 ppm and 136 ppm reveal the lack of symmetry in the fullerene cage in these compounds. The ^{13}C NMR spectra show, for both **3** and **4**, the presence of the fulleropyrrolidine ring on the 6,6-ring junction of the C_{60} framework at ~ 77 ppm and ~ 79 ppm, and those corresponding to the carbons of the fulleropyrrolidine ring appear at ~ 73 ppm (C2) and ~ 70 ppm (C5).

HRMS verified the proposed structure. MALDI-TOF spectrum for compound **3**, registered in the negative mode of detection, shows a peak at $m/z = 1169.2330$, corresponding to the odd-electron molecular ion $\text{M}^{\bullet-}$, while the MALDI-TOF spectrum, in positive mode of detection, for compound **4** shows a peak in $m/z = 1192.2223$ corresponding to $[\text{M} + \text{Na}]^+$. (See Experimental Section and Supporting Information).

In order to determine the spatial arrangement of the protons attached to carbons C2 and C5, NOE experiments were performed on both diastereoisomers **3** and **4**. (See Supporting Information Figure S7 and S19, respectively). In both cases, NOE effect was observed showing the spatial proximity of H2 and H5 protons, indicating the *cis* arrangement in both diastereomers.

With the aim of assigning the absolute stereochemistry of the new stereogenic centers formed in the reaction, namely C2 and C5 of the pyrrolidine ring, we registered the CD spectra of both **3** and **4**, (see Figure 1). The CD spectra do not present an exactly mirror relationship as expected for two diastereomers. However, inspection of the region around 430 nm, considered as the finger print for every fullerene monoadductus, features peaks with opposite sign corresponding to opposite configurations at the new formed stereogenic centers (Figure 1). Indeed, by using the sector rule,¹⁹ proposed for fullerene derivatives,¹³⁻¹⁵ which links the Cotton effect (CE) associated with this UV-vis band and the stereochemical environment around the 6,6 junction, we could assign the configuration of both C2 and C5 atoms.

Compound **3-cis** showed in its CD spectrum a positive CE, which is consistent with a (2*S*,5*R*) stereochemistry (Figure 1, a). The negative CE observed in the CD spectrum of **4-cis**, indicates a (2*R*,5*S*) configuration (Figure 1, b).

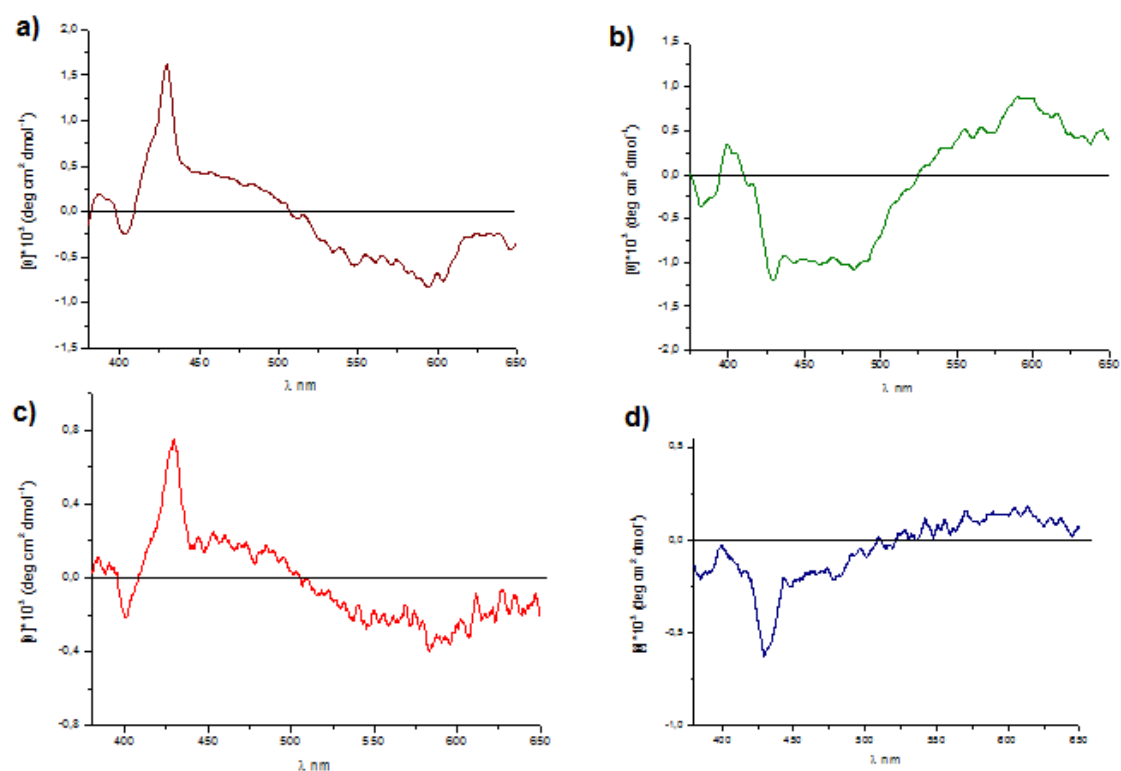


Figure 1. CD spectra of fulleroyrrolidines a) **3**-(2*S*,5*R*)-*cis*, b) **4**-(2*R*,5*S*)-*cis*, c) **5**-(2*R*,5*R*)-*trans*, d) **6**-(2*S*,5*S*)-*trans* in CH₂Cl₂ (conc, 4×10^{-4} M).

The stereochemical outcome stems from the attack of the C₆₀ onto the *Re*,*Si* face of the prochiral C2 and C5 respectively in the case of the stereoisomer **3** and onto the opposite face for the stereoisomer **4** (See Figure 2). The slight diastereomeric excess is consistent with a weak orientation (chiral induction) of the steroid moiety toward the *Re*,*Si* face.

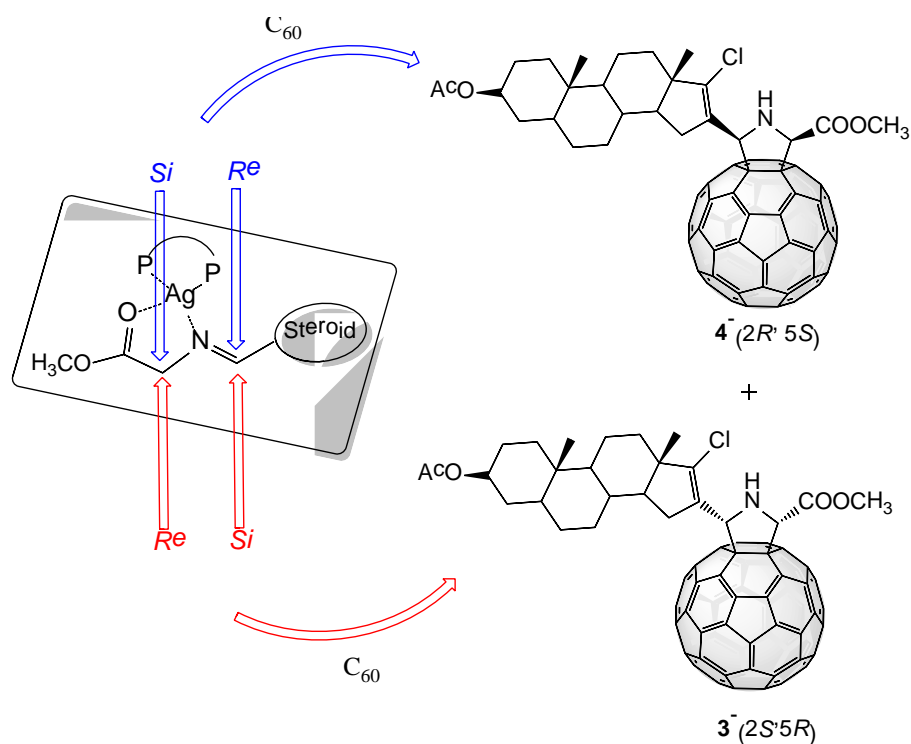


Figure 2. Plausible pathway for the cycloaddition reaction of C_{60} on the chiral metal complex.

In order to ascertain if the previously reported catalytic systems^{15,16} were able to maintain their stereocontrol in the 1,3-dipolar cycloaddition of azomethine ylides onto C_{60} even in the presence of a chiral moiety such as the imine **2**, we performed the reactions using the conditions previously reported. Thus, the catalytic complex Cu(II)–Fesulphos at 0 °C, directs the 1,3-dipolar cycloaddition of steroid iminoester **2** onto C_{60} affording *cis*-(2*S*,5*R*)-2-methoxycarbonyl-5-(3' β -acetoxy-17'-chloro-5' α -16'-androstene) pyrrolidino[3,4:1,2] [60]fullerene (**3**) with an excellent diastereomeric induction. (See Scheme 2 and Table 1, entry 2).

Table 1. Asymmetric Cu(II) and Ag(I)-catalysed 1,3-dipolar cycloadditions of steroid azomethine ylide to C₆₀ using different chiral ligands.

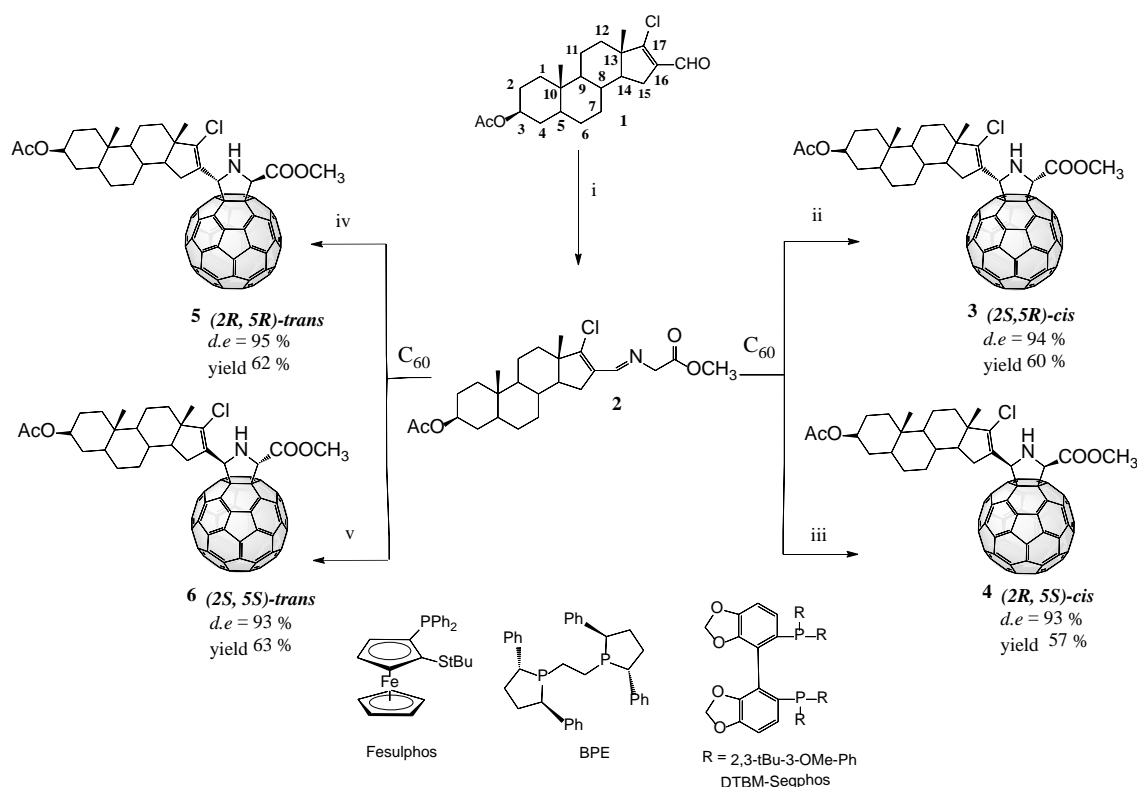
Entry ^a	Metal/Ligand ^a	Temp °C	Conversion % ^b	3 -(2 <i>S</i> ,5 <i>R</i>)- <i>cis</i> %	4 -(2 <i>R</i> ,5 <i>S</i>)- <i>cis</i> %	5 -(2 <i>R</i> ,5 <i>R</i>)- <i>trans</i> %	6 -(2 <i>S</i> ,5 <i>S</i>)- <i>trans</i> %
1	Ag(AcO)/dppe	rt	60	32	25		
2	Cu(AcO) ₂ /FeSulphos	0	80	94	6		
3	AgOAc/(<i>R,R</i>)-BPE	0	88	7	93		
4	Cu(OTf) ₂ / <i>R</i> -DTBM-Segphos/Et ₃ N	rt	90			95	5
5	Cu(OTf) ₂ / <i>S</i> -DTBM-Segphos/Et ₃ N	rt	91			7	93

(a) Conditions (cycloaddition): metal salt (10% mol), chiral ligand (10% mol), anhydrous toluene, 2 h; (b) determined by hplc analysis.

Changing the catalytic system to AgOAc/(*R,R*)-BPE at the same temperature (See Scheme 2 and Table 1, entry 3) the *cis*-diastereoisomer (2*R*,5*S*) (*cis*-2-methoxycarbonyl-5-(3'β-acetoxy-17'-chloro-5'α-16'-androstene)pyrrolidino[3,4:1,2][60]fullerene (**4**) with the opposite configuration is obtained also with excellent diastereoselectivity. Despite the stereoselectivity is slightly lower as result of a mismatched chiral induction between the steroid (2*R*) and the catalyst (2*S*), the system Ag/BPE is still able to maintain high *de* values. In both catalysts, the counterion of the metal salt is acetate because it acts as a base and, probably, occupies the vacancy in the metal coordination sphere, thus enabling a better stereodifferentiation.¹⁵

For completing the diastereodivergent study was important to obtain the *trans*-diastereoisomers. For this, previously reported conditions were used^{16,17} employing the bulky chiral ligand DTBM-Segphos.

In this regard, the complex Cu(II)–(*R*)-DTBM-Segphos in the presence of triethylamine as a base at room temperature gives a *trans*-diastereoisomer **5**, (See Scheme 2 and Table 1, entry 4) with retention time of 5.9 min and a positive CE at 430 nm, corresponding with a (2*R*,5*R*) configuration (See Figure 1, c). The use of Cu(II)–(*S*)-DTBM-Segphos under the same conditions gives a *trans*-diastereoisomer **6** (retention time 8.4 min) with (2*S*,5*S*), (See Scheme 2 and Table 1, entry 5) configuration, consistent with the negative CE at 430 nm, as it is shown in the CD spectrum (Figure 1, d).



Scheme 2. Stereodivergent synthesis of the four diastereoisomers of 2-methoxycarbonyl-5-(3'β-acetoxy-17'-chloro-5'α-16'-androstene)pyrrolidine[3,4:1,2][60]fullerene.

Reagents and conditions: (i) $\text{ClNH}_3\text{CH}_2\text{COOEt}$, Et_3N , DCM anhydrous, rt; (ii) $\text{Cu}(\text{AcO})_2$, Fesulphos, 0 °C, 2 h; (iii) $\text{Ag}(\text{OAc})$, BPE, 0 °C, 2 h; (iv) $\text{Cu}(\text{OTf})_2$, *R*-DTBM-Segphos, Et_3N , 2 h, rt; (v) $\text{Cu}(\text{OTf})_2$, *S*-DTBM-Segphos, Et_3N , 2 h, rt.

To confirm the *trans* stereochemistry of diastereoisomers **5** and **6**, ROESY-1D spectra were recorded and no interaction between H2 and H5 protons was observed, indicating the *trans* arrangement between them. (See Supporting Information, Figure S30 for **5** and Figure S41 for **6**).

The spectroscopical data confirm the structure of both *trans*-diastereomeric hybrids **5** and **6** which are very similar to the *cis*-diastereoisomers **3** and **4**. (see the Experimental Section and Supporting Information). ^1H NMR shows the signals corresponding to the protons of the pyrrolidine ring at 5.57 ppm (H2) and 5.84 ppm (H5) for **5** and 5.57 ppm (H2) and 5.81 ppm (H5) for **6**.

The ^{13}C NMR spectra shows, for both **5** and **6**, signals at ~76 ppm and ~79 ppm, indicating the 6,6-ring junction of C_{60} to pyrrolidine ring, and those corresponding to the carbons of the fulleropyrrolidine ring which appear at ~74 ppm (C2) and ~70 ppm (C5). Also the signals at the interval of 156-136 ppm, denote the presence of the fullerene cage.

MALDI-TOF spectrum for compound **5**, registered in the positive mode of detection, shows a peak at $m/z = 1170.2405$, corresponding to $[\text{M} + \text{H}]^+$ ion, while the mass spectrum

of **6** shows a peak at $m/z = 1170.2453$ corresponding to $[M + H]^+$ (See Supporting Information).

The chemical structure of all final compounds (**3-6**) was confirmed by combined NMR spectroscopic data from ^1H , ^{13}C , COSY, DEPT, HMQC, and HMBC experiments. (See Supporting Information).

Also the specific optical rotation measurements for the hybrids were carried out. Thus, the experimentally determined values for $[\alpha]_{\text{D}}^{20}$ were: $+125^\circ$ for **3-cis**, $+40^\circ$ for **4-cis**, $+105^\circ$ for **5-trans** and $+35^\circ$ for **6-trans**.

The 1,3-dipolar cycloaddition between the steroidal imine **2** and [60]fullerene is coherent with the previous results obtained by Martin *et al.* for the synthesis of stereodivergent chiral fulleropyrrolidines.¹⁵

CONCLUSIONS

In summary, we have carried out for the first time a controlled diastereoselective synthesis of chiral steroid-fulleropyrrolidines by reaction of copper or silver *N*-metalated azomethine ylide and chiral ligands. Depending on the chiral metal/ligand employed, it is possible to obtain the pairs of *cis* and *trans* diastereomers with good diastereomeric excesses and good yields. The new steroid-fullerene hybrids have been characterized by spectroscopic and dichroic study which reveal the formation of the four different stereoisomers. Importantly, these results show that the azomethine ylides stabilized by an allylic group generated from the steroidal imine **2**, behaves in a similar favorable way to that observed for the benzyl related compounds in previous studies (see above).

In contrast to previous papers involving steroid-fullerene hybrids where only poor stereoselectivities were achieved, the stereodivergent protocol now presented shows the efficiency for the preparation of diastereoselective derivatives in a controlled manner, thus paving the way to a great variety of optically active fullerene derivatives.

EXPERIMENTAL

General Experimental Methods. All reactions were performed using an atmosphere of argon and oven-dried glassware. Solvents were treated prior to use according to the standard methods. The commercially available reagents were used without further purification. Reactions were monitored by thin-layer chromatography carried out on 0.25 mm silicagel plates (230–400 mesh). Flash column chromatography was performed using

silica gel (60 Å, 32–63 µm). FTIR spectra were recorded in CHCl₃. ¹H-NMR spectra were recorded at 700 MHz and ¹³C-NMR at 175 MHz; the one-bond heteronuclear correlation (HMQC) and the long-range ¹H–¹³C correlation (HMBC) spectra were obtained by use of the inv4gs and the inv4gslplrnd programs. All MS-ESI and HRMS-MALDI (dithranol as matrix) experiments were carried out in negative and positive modes of detection. UV/vis spectra were recorded in CHCl₃. Microanalysis was performed with a CHN instrument. Optical rotations were measured using a polarimeter with a thermally jacketed 10 cm cell at 20°C (concentration given as g/100ml). The CD spectra were recorded in dichloromethane. A high-performance liquid chromatography (HPLC) system (column dimensions, 4.6 × 250 mm; flow rate 1.0 mL min⁻¹, injection volume 15 µL, eluent toluene:acetonitrile 9:1) was used to determine the purity of the compounds synthesized and *de* values. All these values were monitored in a 320 nm spectrophotometer detector.

***N-methyl [(3β-acetoxy-17-chloro-16-androstene-16-yl) methylene] glycinate* (2).** A mixture of glycine methyl ester hydrochloride 75 mg (0.6 mmol) and triethyl amine 0.26 mL (1.8 mmol) with molecular sieves (4Å) in 40 mL of anhydrous dichloromethane (DCM) was stirred for one hour. Then, 3β-acetoxy-17-chloro-16-formyl-5α-16-androstene 160 mg (0.42 mmol) was added and the reaction was stirred overnight at room temperature. The reaction mixture was filtered to separate the molecular sieves and the filtrate was washed twice with water and subsequently over anhydrous MgSO₄. The organic extract was concentrated, obtaining a yellow oil. Yield 77 % (145 mg). ¹H-NMR (700 MHz, CDCl₃, δ ppm): 8.22 (s, 1H, CH=N), 4.67 (m, 1H, H3), 4.31 (s, 2H, NCH₂CO), 3.76 (s, 3H, CH₃O), 2.63 (dd, *J* = 14.7 Hz, *J* = 6.0 Hz, 1H, H15), 2.04 (m, 1H, H15), 2.01 (s, 3H, CH₃-CO), 1.84 (m, 1H, H2), 1.78 (m, 1H, H7), 1.73 (m, 1H, H1), 1.67 (m, 2H, H11), 1.63 (m, 1H, H4), 1.59 (m, 1H, H14), 1.57 (m, 1H, H8), 1.51 (m, 1H, H2), 1.46 (m, 1H, H12), 1.42 (m, 1H, H12), 1.38 (m, 1H, H4), 1.33 (m, 2H, H11), 1.30 (m, 2H, H6), 1.20 (m, 1H, H5), 1.03 (m, 1H, H1), 0.97 (m, 1H, H7), 0.84 (m, 1H, H9), 0.82 (s, 3H, CH₃-19), 0.74 (s, 3H, CH₃-18). ¹³C-NMR (175 MHz, CDCl₃, δ ppm): 170.7 (C=O), 170.6 (C=O), 160.7 (CH=N), 152.7 (C17), 134.9 (C16), 73.5 (C3), 62.3 (NCH₂CO), 54.6 (C14), 53.9 (C9), 52.2 (CH₃O), 49.9 (C13), 44.8 (C5), 36.5 (C1), 35.7 (C10), 34.0 (C8), 33.9 (C12), 33.4 (C4), 31.1 (C7), 28.3 (C15), 28.2 (C6), 27.4 (C2), 21.5 (CH₃-CO), 20.7 (C11), 15.2 (C18), 12.2 (C19). IR (CHCl₃): ν = 3242, 2927, 2858, 1732 (C=O), 1674, 1244, 1026, 756 cm⁻¹. C₂₅H₃₆ClNO₄ (449.23): calcd. C 66.72, H 8.06; found C 66.75, H 8.10. MS (MALDI-TOF) *m/z*: [M + Na]⁺ 472.3.

cis-(2S,5R)-2-Methoxycarbonyl-5-(3'β-acetoxy-17'-chloro-5'α-16'-androstene)pyrrolidino[3,4:1,2][60]fullerene (3). A mixture of Cu(OAc)₂ (1 mg) and Fesulfos (2.3 mg) in anhydrous toluene (10 mL) was stirred for one hour at room temperature. Then, 20 mg of steroid α-iminoester **1** was added and the yellow mixture was cooled at 0°C. Finally, 30 mg (0.042 mmol) of C₆₀ was also added. The reaction mixture was stirred for two hours, and afterwards, it was quenched with an aqueous saturated ammonium chloride solution (3 mL). The mixture was extracted with toluene (3 x 1mL), and the combined extracts were washed with brine (3 mL). The organic layer was dried over MgSO₄ and concentrated *in vacuo*. The solid crude was purified by column chromatography on silica gel, using CS₂ to elute unreacted C₆₀ and dichloromethane to elute the corresponding pyrrolidine [3,4:1,2][60]fullerene **3**. Brown solid, melting point over 350° C. Chemical yield: 60 % (29 mg). HPLC: toluene, flow rate 1 mL/min, *t_R* = 6.96 min. [α]_D²⁰ = +125° (c 2 x 10⁻⁴ CH₂Cl₂). ¹H-RMN (700 MHz, CDCl₃, δ ppm): 5.81 (s, 1H, H5), 5.57 (s, 1H, H2), 4.65 (m, 1H, H3'), 3.90 (s, 3H, CH₃O), 2.60 (dd, *J* = 14.5 Hz, *J* = 6.3 Hz, 1H, H15'), 2.44 (dd, *J* = 14.4 Hz, *J* = 11.6 Hz, 1H, H15'), 2.03 (s, 3H, CH₃CO), 1.80 (m, 1H, H2'), 1.77 (m, 1H, H12'), 1.74 (m, 1H, H7'), 1.68 (m, 1H, H1'), 1.61 (m, 1H, H8'), 1.60 (m, 1H, H11'), 1.58 (m, 1H, H4'), 1.47 (m, 1H, H2'), 1.37 (m, 1H, H11'), 1.35 (m, 1H, H4'), 1.33 (m, 1H, H14'), 1.30 (m, 2H, H6'), 1.21 (m, 1H, H12'), 1.17 (m, 1H, H5'), 1.01 (s, 3H, CH₃-18'), 0.99 (m, 1H, H1'), 0.86 (s, 3H, CH₃-19'), 0.82 (m, 1H, H7'), 0.67 (m, *J* = 4.6 Hz, 1H, H9'). ¹³C-RMN (175 MHz, CDCl₃, δ ppm): 170.7 (C=O); 169.9 (C=O-C2), 153.3, 152.5, 151.7, 150.7, 147.2, 147.0, 146.5, 146.4, 146.33, 146.29, 146.2, 146.1, 145.9, 145.5, 145.4, 145.38, 145.35, 145.3, 145.2, 145.16, 145.1, 144.9 (C17'), 144.5, 144.3, 144.2, 144.1, 143.2, 143.1, 143.0, 142.8, 142.7, 142.7, 142.5, 142.4, 142.3, 142., 142.1, 142.0, 141.9, 141.8, 141.6, 140.0, 139.8, 139.7, 139.6, 136.6, 136.1, 135.3, 134.8, 132.0 (C16'), 79.8 (Csp³ C₆₀), 76.9 (Csp³ C₆₀), 74.0 (C2), 73.5 (C3'), 70.5 (C5), 55.2 (C14'), 54.5 (C9'), 53.5 (CH₃O), 48.9 (C13'), 44.6 (C5'), 36.3 (C1'), 35.7 (C10'), 34.2 (C12'), 33.9 (C4'), 33.7 (C8'), 32.2 (C15'), 31.4 (C7'), 28.2 (C6'), 27.4 (C2'), 21.5 (CH₃CO), 20.8 (C11'), 15.1 (C18'), 12.2 (C19'). IR (CHCl₃): ν = 2924, 2854, 1737 (C=O), 1249, 1025, 755 cm⁻¹. HRMS (MALDI-TOF) *m/z*: [M^{•+}] Calcd for C₈₅H₃₆ClNO₄ 1169.2338; Found 1169.2330.

cis-(2R,5S)-2-Methoxycarbonyl-5-(3'β-acetoxy-17'-chloro-5'α-16'-androstene)pyrrolidino[3,4:1,2][60]fullerene. (4). A mixture of AgOAc (1 mg) and BPE (2.3 mg) in toluene anhydrous (10 mL) in toluene anhydrous (10 mL) was stirred for one hour at room

temperature. Then, 20 mg of steroid α -iminoester **1** was added and the yellow mixture was cooled at 0°C. Finally, 30 mg (0.042 mmol) of C₆₀ was also added. The reaction mixture was stirred for two hours, and afterwards, it was quenched with an aqueous saturated ammonium chloride solution (3 mL). The mixture was extracted with toluene (3 x 1 mL), and the combined extracts were washed with brine (3 mL). The organic layer was dried over MgSO₄ and concentrated *in vacuo*. The solid crude was purified by column chromatography on silica gel, using CS₂ to elute unreacted C₆₀ and dichloromethane:ethyl acetate (100:1) to elute the corresponding pyrrolidine [3,4:1.2][60]fullerene **4**. Brown solid, melting point over 350° C. Chemical yield: 57 % (27 mg). HPLC: toluene, flow rate 1 mL/min, t_R = 10.71 min. $[\alpha]_D^{20}$ = +40° (c 2 x 10⁻⁴ CH₂Cl₂). ¹H-RMN (700 MHz, CDCl₃, δ ppm): 5.86 (s, 1H, H5), 5.58 (s, 1H, H2), 4.71 (m, 1H, H3'), 3.90 (s, 3H, CH₃O), 2.72 (dd, J = 14.5 Hz, J = 6.4 Hz, 1H, H15'), 2.14 (dd, J = 13.8 Hz, J = 11.9 Hz, 1H, H15'), 2.03 (s, 3H, CH₃CO), 1.83 (m, 1H, H2'), 1.80 (m, 1H, H7'), 1.77 (m, 1H, H12'), 1.73 (m, 1H, H14'), 1.70 (m, 1H, H1'), 1.66 (m, 1H, H11'), 1.62 (m, 1H, H8'), 1.57 (m, 1H, H4'), 1.49 (m, 1H, H2'), 1.40 (m, 1H, H12'), 1.38 (m, 1H, H4'), 1.35 (m, 1H, H11'), 1.33 (m, 1H, H6'), 1.29 (m, 1H, H6'), 1.24 (m, 1H, H5'), 1.09 (m, 1H, H7'), 1.03 (m, 1H, H1'), 0.84 (m, 1H, H9'), 0.83 (s, 3H, CH₃-19'), 0.75 (s, 3H, CH₃-18'). ¹³C-RMN (175 MHz, CDCl₃, δ ppm): 170.9 (CO-20'), 170.1 (CO-C2), 153.6, 152.9, 152.1, 150.9, 147.3, 147.2, 146.7, 146.5, 146.44, 146.42, 146.38, 146.2, 146.1, 146.0, 145.7, 145.60, 145.57 (C17'), 145.5, 145.47, 145.39, 145.35, 145.33, 145.30, 144.5, 144.48, 144.45, 144.3, 143.27, 143.25, 143.17, 142.92, 142.89, 142.84, 142.81, 142.76, 142.6, 142.4, 142.3, 142.2, 142.14, 142.11, 142.09, 142.06, 141.98, 141.2, 139.90, 139.83, 139.75, 136.8, 136.25, 136.17, 135.0, 131.7 (C16'), 79.4 (Csp³ C₆₀), 76.9 (Csp³ C₆₀), 73.8 (C2), 73.7 (C3'), 70.7 (C5), 55.12 (C14'), 54.7 (C9'), 53.0 (OCH₃), 49.1 (C13'), 44.9 (C5'), 36.6 (C1'), 35.9 (C10'), 34.10 (C8'), 34.08 (C4'), 33.6 (C12'), 32.1 (C15'), 31.4 (C7'), 28.4 (C6'), 27.5 (C2'), 21.6 (C21'), 20.8 (C11'), 16.2 (C18'), 12.3 (C19'). IR (CHCl₃): ν = 2928, 2853, 1737 (C=O), 1249, 1025, 754 cm⁻¹. HRMS (MALDI-TOF) m/z : [M + Na]⁺ Calcd for C₈₅H₃₆ClNO₄Na 1192.2225; Found 1192.2223.

***trans*-(2*R*,5*R*)-2-Methoxycarbonyl-5-(3' β -acetoxyl-17'-chloro-5' α -16'-androstene)**

***pyrrolidino*[3,4:1,2][60]fullerene. (5).** A mixture of cooper(II)triflate (1.9 mg) and *R*-DTBM-Segphos (6 mg) in toluene anhydrous (10 mL) was stirred for one hour at room temperature, getting a solution with a slight blue colour. Then, 23.5 mg of steroid α -iminoester **1** was added obtaining a yellow mixture. Finally, C₆₀ (33.6 mg) and 0.2 mL of

triethylamine were added. The reaction mixture was stirred for two hours, and afterwards, it was quenched with an aqueous saturated ammonium chloride solution (5 mL). The mixture was extracted with toluene (3 x 10 mL), and the combined extracts were washed with brine (10 mL). The organic layer was dried over MgSO₄ and concentrated in vacuo. The crude product was purified by column chromatography on silica gel, using CS₂ to elute unreacted C₆₀ and dichloromethane to elute the corresponding pyrrolidine [3,4:1,2][60]fullerene **5**. Chemical yield: 62 % (29 mg). Brown Solid. HPLC: toluene, flow rate 1 mL/min, t_R = 5.91 min. $[\alpha]_D^{20}$ = +105° (c 2 x 10⁻⁴ CH₂Cl₂). ¹H-NMR(700 MHz, CDCl₃, δ ppm): 5.84 (s, 1H, H5), 5.57 (s, 1H, H2), 4.65 (m, 1H, H3'), 3.66 (s, 3H, CH₃O), 2.60 (m, 1H, H15'), 2.44 (m, 1H, H15'), 2.01 (s, 3H, CH₃CO), 1.82 (m, 1H, H2'), 1.78 (m, 1H, H12'), 1.76 (m, 1H, H7'), 1.70 (m, 1H, H1'), 1.63 (m, 1H, H11'), 1.61 (m, 1H, H8'), 1.58 (m, 1H, H4'), 1.50 (m, 1H, H2'), 1.39 (m, 1H, H11'), 1.38 (m, 1H, H4'), 1.37 (m, 1H, H14'), 1.33 (m, 1H, H6'), 1.29 (m, 1H, H6'), 1.23 (m, 1H, H12'), 1.18 (m, 1H, H5'), 1.02 (m, 1H, H1'), 0.89 (s, 3H, CH₃-18'), 0.85 (s, 3H, CH₃-19'), 0.82 (m, 1H, H7'), 0.68 (m, 1H, H9'). ¹³C-NMR (175 MHz, CDCl₃, δ ppm): 170.61 (CO-20'), 169.70 (CO-C2), 147.14, 147.00, 146.34, 146.23, 145.99, 145.94, 145.57, 145.44, 145.40, 145.33, 145.26, 145.14, 144.51, 143.31, 144.25, 143.15, 143.09, 142.74, 142.73, 142.68, 142.65, 142.54, 142.41, 142.30, 142.28, 142.14, 142.05, 142.04, 142.00, 141.91, 140.03, 139.68, 137.86 (C17'), 129.02 (C16'), 79.68 (Csp³ C₆₀), 76.38 (Csp³ C₆₀), 74.09 (C2), 73.47 (C3'), 70.74 (C5), 70.57 (OCH₃), 55.17 (C14'), 54.48 (C9'), 48.86 (C13'), 44.64 (C5'), 36.32 (C1'), 35.71 (C10'), 34.25 (C4'), 33.90 (C12'), 33.71 (C8'), 31.34(C15'), 29.69 (C7'), 28.21 (C6'), 27.36 (C2'), 21.43 (C21'), 20.66 (C11'), 14.35 (C18'), 12.15 (C19'). IR (CHCl₃): ν = 2923, 2854, 1738, 1678, 1522, 743 cm⁻¹. HRMS (MALDI-TOF) m/z: [M + H]⁺ Calcd for C₈₅H₃₇ClINO₄ 1170.2411; Found 1170.2405.

trans-(2S,5S)-2-methoxycarbonyl-5-(3'β-acetoxy-17'-chloro-5'α-16'-androstene

pyrrolidino[3,4:1,2][60]fullerene (6). A mixture of cooper(II)triflate (2 mg) and S-DTBM-Segphos (6 mg) in toluene anhydrous (10 mL) was stirred for one hour at room temperature, getting a solution with a slight blue colour. Then, 23.5 mg of steroid α-iminoester **1** was added obtaining a yellow mixture. Finally, C₆₀ (33 mg) and 0.2 mL of triethylamine were added. The reaction mixture was stirred for two hours, and afterwards, it was quenched with an aqueous saturated ammonium chloride solution (5 mL). The mixture was extracted with toluene (3 x 10mL), and the combined extracts were washed with brine (10 mL). The organic layer was dried over MgSO₄ and concentrated in vacuo.

The crude product was purified by column chromatography on silica gel, using CS₂ to elute unreacted C₆₀ and dichloromethane:ethyl acetate (99:1) to elute the corresponding pyrrolidine [3,4:1.2][60]fullerene **6**. Chemical yield: 63 % (30 mg). Brown Solid. HPLC: solvent toluene, flow rate 1 mL/min, *t*_R = 8.42 min. [α]_D²⁰ = +35° (c 2 x 10⁻⁴ CH₂Cl₂). ¹H-NMR (700 MHz, CDCl₃, δ ppm): 5.81 (s, 1H, H5), 5.57 (s, 1H, H2), 4.69 (m, 1H, H3'), 3.90 (s, 3H, CH₃O), 2.60 (m, 1H, H15'), 2.44 (m, 1H, H15'), 2.01 (s, 3H, CH₃CO), 1.82 (m, 1H, H2'), 1.77 (m, 1H, H12'), 1.76 (m, 1H, H7'), 1.70 (m, 1H, H1'), 1.61 (m, 1H, H11'), 1.60 (m, 1H, H8'), 1.58 (m, 1H, H4'), 1.49 (m, 1H, H2'), 1.39 (m, 1H, H4'), 1.38 (m, 1H, H14'), 1.37 (m, 1H, H11'), 1.34 (m, 1H, H6'), 1.31 (m, 1H, H6'), 1.22 (m, 1H, H12'), 1.18 (m, 1H, H5'), 1.02 (m, 1H, H1'), 0.89 (s, 3H, CH₃-18'), 0.86 (s, 3H, CH₃-19'), 0.83 (m, 1H, H7'), 0.67 (m, 1H, H9'). ¹³C-NMR (175 MHz, CDCl₃, δ ppm): 170.7 (CO-20'), 169.9 (CO-C2), 147.1, 147.0, 146.3, 146.0, 145.6, 145.4, 145.3, 145.2, 144.5, 144.3, 144.2, 143.2, 143.1, 142.74, 142.67, 142.65, 142.5, 142.4, 142.3, 142.1, 142.05, 142.00, 141.9, 141.8, 140.04, 140.03, 139.7, 138.0 (C17'), 128.9 (C16'), 79.8 (Csp³ C₆₀), 77.2 (Csp³ C₆₀), 74.1 (C2), 73.5 (C3'), 70.6 (OCH₃), 70.5 (C5), 55.2 (C14'), 54.5 (C9'), 48.9 (C13'), 44.6 (C5'), 36.4 (C1'), 35.7 (C10'), 34.3 (C4'), 33.9 (C12'), 33.7 (C8'), 31.4 (C15'), 29.4 (C7'), 28.2 (C6'), 27.4 (C2'), 21.5 (C21'), 20.7 (C11'), 14.2 (C18'), 12.2 (C19'). IR (CHCl₃): ν = 2924, 2853, 1738, 1250, 1025, 723 cm⁻¹. HRMS (MALDI-TOF) *m/z*: [M + H]⁺ Calcd for C₈₅H₃₇ClINO₄ 1170.2411; Found 1170.2453.

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SUPPORTING INFORMATION

The Supporting Information is available free of charge on the ACS Publications website:

¹H-NMR and ¹³C NMR (1D and 2D), MS, IR and UV spectra and HPLC chromatograms.

REFERENCES

- (1) (a) Huang, Y. Y.; Sharma, S.K.; Yin, R.; Agrawal, T.; Chiang, L.Y.; Hamblin, M.R. *J. Biomed. Nanotechnol.* **2014**, *10*, 1918–1936. (b) Partha, R.; Conyers, J. L. *Int. J. Nanomed.* **2009**, *4*, 261-271.
- (2) Mehta, G.; Singh V. *Chem. Soc. Rev.* **2002**, *31*, 324-334.
- (3) Salunke, D. B.; Hazra, B. G.; Pore, V. S. *Curr. Med. Chem.* **2006**, *13*, 813–847.
- (4) Burendic, E.; Penov-Gasi, K.; Medic-Mejacevic, L. *Hemijski Pregled*, **2002**, *43*, 82-84.
- (5) Fong, R. II; Schuster, D. I.; Wilson, S. R. *Org. Lett.* **1999**, *1*, 729-732.
- (6) Ishi, I, T.; Shinkai, S., *Tetrahedron* **1999**, *55*, 12515-12530.
- (7) Li, L.; Hu, Y.; Wu, Y.; Wu, Y.; Yue, J.; Yang, F., *J. Chem. Soc., Perkin Trans. 1*, **2001**, 617-621.
- (8) Bjelakovic, M. S.; Godjevac, D. M.; Milic, D. R. *Carbon*, **2007**, *45*, 2260-2265.
- (9) MacFarland, D.; Zhang, J.; Zhou, Z.; Lenk, R. P.; Wilson, S. R. U.S. Pat. Appl. Publ. **2008**, US 20080214514 A1 20080904.
- (10) Bjelakovic, M.; Kop, T.; Baosic, R.; Zlatovic, M.; Zekic, A.; Maslak, V.; Milic, D. *Monatsh. Chem.* **2014**, *145*, 1715-1725.
- (11) Bjelakovic, M. S.; Kop, T. J.; Vljajic, M.; Djordjevic, J.; Milic, D. R. *Tetrahedron* **2014**, *70*, 8564-8570.
- (12) Coro, J.; Rodriguez, H.; Rivera, D. G.; Suarez, M.; Molero, D.; Herranz, M. A.; Martinez-Alvarez, R.; Filippone, S.; Martin, N. *Eur. J. Org. Chem.* **2009**, 4810-4817.
- (13) Ruiz, A.; Coro, J.; Almagro, L.; Ruiz, J. A.; Molero, D.; Maroto, E. E.; Filippone, S.; Herranz, M. A.; Martinez-Alvarez, R.; Sancho-Garcia, J. C.; Di Meo F.; Suarez, M.; Martín, N. *J. Org. Chem.* **2013**, *78*, 2819-2826.
- (14) Ruiz, A.; Morera-Boado, C.; Almagro, L.; Coro, J.; Maroto, E. E.; Herranz, M. A.; Filippone, S.; Molero, D.; Martinez-Alvarez, R.; Garcia de la Vega, J. M.; Suárez, M.; Martín, N. *J. Org. Chem.* **2014**, *79*, 3473-3486.
- (15) Filippone, S.; Maroto, E. E.; Martín-Domenech, A.; Suárez, M.; Martín, N. *Nat. Chem.* **2009**, *1*, 578–582.
- (16) Maroto, E. E.; Filippone, S.; Martín-Domenech, A.; Suárez, M.; Martín, N. *J. Am. Chem. Soc.* **2012**, *134*, 12936–12938.
- (17) Maroto, E. E.; Filippone, S.; Suárez, M.; Martínez-Álvarez, R.; de Cózar, A.; Cossío, F. P.; Martín, N. *J. Am. Chem. Soc.* **2014**, *136*, 705–712.

- (18) Maroto, E. E.; Izquierdo, M.; Reboredo, S.; Marco-Martínez, J.; Filippone, S.; Martín, N. *Acc. Chem. Res.* **2014**, *47*, 2660–2670.
- (19) Wilson, S. R.; Lu, Q.; Cao, J.; Wu, Y.; Welch, C. J.; Schuster, D. I. *Tetrahedron* **1996**, *52*, 5131–5142.